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Design and synthesis of a new homologues series of 2-oxo-

benzoazepinone derivatives as potent estrogen receptor alpha inhibitors

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Abstract

Selective estrogen receptor modulators (SERMs) bind to estrogen receptors and prevent estrogen from acting on breast cancer cells. A SERM prevents estrogen from attaching to the cancer cell and preventing the cancer cell from receiving signals from estrogen that would otherwise cause it to grow and multiply. The objective of this study is to create innovative Benzoazepinone derivatives that replicate the SERMs' capacity to attach to estrogen receptor protein in the form of tiny, drug-like molecules. (S)-1amino-3-methyl-1,3,4,5-tetrahydro-2H-benzo[d]azepin-2-one undergoes amidation with (tert-butoxycarbonyl)-L-alanine, gives tert-butyl ((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl)carbamate (Inter-1), which is further undergoing deprotection gives (S)-2-amino-N-((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1vl)propanamide (Inter-2) (Figure 1). Inter-2 on reaction with different substituted aromatic carboxylic acids and yields N-((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl) substituted benzamide derivatives (T16-T30) with the formation of amide bond. All the compounds were screened for their *in-vitro* cytotoxicity activity using Vero and MDA MB 231 cell lines by MTT assay. IC50 values from Cytotoxicity studies by MTT assay ranges from 15.105µg/ml to 155µg/ml. A total of 15 compounds were synthesized by using a diverse scheme and the title compounds have exhibited low to high in-vitro anticancer activity with MDA MB 231 cells. Compared to the standard (Raloxifene 6 µg/ml), the developed compounds T22 (15.105 µg/ml), T24 (37.206 µg/ml), T18 (41.200 µg/ml), T20 (58.302 µg/ml). Finally, four compounds might be used as a lead molecule for future development into a therapeutically viable anti-ER positive breast cancer drug from the 2-oxobenzoazepinone derivatives family.

Keywords: ER alpha inhibitors, Raloxifene, In vitro, MDA MB-231, Bezoazepenine.

 Full length article
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1. Introduction

The most fatal disease is cancer, and efforts are being made to find possible anticancer medications. One of the most prevalent cancers afflicting women worldwide, both in industrialised and developing nations, is breast cancer [1]. 14% of all female malignancies in India are breast cancers. Breast cancer was recorded as 1,62,468 new cases and 87, 090 fatalities in India in 2021, according to Globocan statistics [2]. High mortality is influenced by a lack of accurate diagnosis and expensive medical care. Estrogen Receptor (ER) positivity accounts for two-thirds of all breast cancers, hence efforts are being made to identify patients who would benefit from antiestrogen medication. Breast cancer's dependence on oestrogen is a distinctive trait of the illness that can be targeted to successfully restrict growth and/or halt tumour formation. The current approach for treating hormone-dependent breast cancer is, in fact, to stop oestrogen from acting on tumour cells in one of three ways: (a) by preventing oestrogen from binding to its primary target ER with an antiestrogen like tamoxifen/raloxifen [3-8]; (b) by preventing oestrogen synthesis with an aromatase inhibitor [9–15]; or (c) by reducing ER protein levels with a pure antiestrogen like fulvestrant. The emergence of resistance, however, frequently limits the efficacy of these modern endocrine therapies.

1.1 Drugs existing for ER-positive breast cancer therapy and drawbacks

Example of SERMs (Selective Estrogen Receptor Modulators) The effects of oestrogen on breast tissue are blocked by the drugs Tamoxifen (1) and Raloxifene (2). The oestrogen receptors in breast cells are where SERMs do their action. Estrogen cannot connect to the cell if a SERM is present in the oestrogen receptor because there is no place for it. The signals from oestrogen to grow and multiply are not received by a breast cell if oestrogen is not present. Other bodily tissues, such as the uterus and bones, include cells with oestrogen receptors as well. But based on the type of cell it is in, each oestrogen receptor has a somewhat different shape. As a result, oestrogen receptors on breast cells are distinct from oestrogen receptors on bone cells, and both oestrogen receptors are distinct from oestrogen receptors on uterine cells. Because SERMs are "selective," as their name implies, they can enhance the activity of oestrogen in other cells, including bone, liver, and uterine cells, even while they suppress estrogen's effect in breast cells. Serious adverse reactions from SERMs can include endometrial cancer, stroke, and blood clots. Fulvestrant (3), for instance, is a kind of medication that binds to the ER and, in the course of doing so, causes the ER to be destroyed and downregulated (a selective oestrogen receptor degrader or down regulator, or SERD) [21-22]. Along with older medications like selective oestrogen receptor modulators (SERMs) and aromatase inhibitors, they are used to treat oestrogen receptor-sensitive or progesterone receptor-sensitive breast cancer [23-24]. Candidates for antiestrogen treatment are found since ER positive breast cancer accounts for two thirds of all cases. Breast cancer is a rare illness that can be efficiently managed to reduce growth and/or prevent tumour formation due to its dependence on oestrogen. In fact, the current approach for treating hormone-dependent breast cancer is to prevent oestrogen from acting on tumour cells in one of three ways: (a) by preventing oestrogen from binding to its primary target ER with an antiestrogen like tamoxifen/raloxifene [25]; (b) by preventing oestrogen synthesis with an aromatase inhibitor [23,26-29]; or (c) by lowering ER protein levels with a pure antiestrogen like ful The emergence of resistance, however, frequently limits the efficacy of these modern endocrine The objective of this study is to create medications. innovative Benzoazepinone derivatives that replicate the SERMs' capacity to attach to oestrogen receptor protein in the form of tiny, drug-like molecules. These Benzoazepinone derivatives may help in breast cancer therapy and/or prevention.

2. Experimental Section

2.1 Chemistry

Chemicals used were of the reagent category and purified as needed. Melting points were determined by Lab India digital melting point apparatus. Shimadzu's FTIR spectrometer model was used for recording the IR spectrum of compounds. Bruker DRX-300 spectrometer was used for the determination of NMR spectra in the DMSO solvent with internal standard as a TMS. Shimadzu LCMS 2010A spectrometer was used to examine the MASS spectra of compounds. Responses were checked by thin-layer chromatography (TLC).

2.2 Step 1: Preparation of tert-butyl ((S)-1-(((S)-3-methyl-2oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1-yl)amino)-1oxopropan-2-yl)carbamate (Inter-1)

To a mixture of N-Boc Alanine, HOBt hydrate, MC07JC1876 in THF under nitrogen atmosphere was added N,N Diisopropyl ethyl amine and EDC. The reaction mixture was stirred for 16 h at RT. After completion of the reaction, the solvent was removed under vacuum, the residue taken in ethyl acetate and water, washed with saturated NaHCO₃ solution, 1N HCl, brine solution, dried over sodium sulfate, filtered and concentrated under vacuum to afford pure 10 g of inter-1 in 68% yield. TLC System, 50% Ethyl acetate in hexane Rf Value-0.4.

2.3 Step 2: Preparation of (S)-2-amino-N-((S)-3-methyl-2oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1 yl) propenamide (Inter-2)

2.3.1 Procedure

A stream of anhydrous HCl gas was passed through a stirred solution of inter-1 in dioxane, chilled in ice bath to about 10°C under nitrogen for 10-15 minutes. The solution was capped, then cooling bath removed, it was allowed to warm to RT with stirring for 2-8 h. After completion of the reaction, it was concentrated to afford 6 g of inter-2 in 83.3% yield. TLC System 10% Methanol in chloroform. Rf Value is 0.2.

2.4 Step 3: Preparation of Final Derivatives N-((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1yl)amino)-1-oxopropan-2-yl) substituted benzamide derivatives (T16-T30)

To a mixture of inter-2, HOBt hydrate, (S)-2-hydroxy-3-methyl butyric acid in THF under nitrogen atmosphere was added N,N-diisopropylethylamine, followed by EDC and stirred for 16 h at RT. After completion of the reaction, the solvent was removed under vacuum. The residue was taken in ethyl acetate and water, washed with saturated NaHCO₃ solution, 1 N HCl, brine, dried over sodium sulfate and removed the solvent under vacuum to afford the 4.5 g of MC07JC1990 in 54.8% yield. TLC System, 50% Ethyl acetate in hexane. Rf Value is 0.3. to afford benzoazepinone derivatives T1-T15 in 60-70% yield.

2.5 Pharmacology

2.5.1 Cytotoxicity Screening

The cell culture was centrifuged and the cell count was adjusted to 1.0*10⁵ cells/ml using DMEM medium containing 10% FBS. To each well of a 96 well flat bottom microtitre plate, 100 µl of the diluted cell suspension (approximately 10,000 cells/well) was added. After 24 hours, when the cell population was found adequate, the cells were suspended with 100 µl of different test sample concentrations prepared in maintenance media. The plates were then incubated at 37 °C for 48 h in a 5% CO₂ atmosphere, and microscopic examination was carried out and observations recorded every 24 h. After 48 h, 20 µl of MTT (2mg/ml) in MEM-PR (MEM without Phenol Red) was added. The plates were gently shaken and incubated for 2 h at 37 °C in a 5% CO₂ atmosphere ^[36-40]. The 100µl of DMSO was added and the plates were gently shaken to solubilize the formed formazan. The absorbance was measured using a microplate reader at a wavelength of 540 nm. The percentage cell viability was calculated using the following formula and the concentration of drug or test samples needed to inhibit cell growth by 50% values were granted from the dose-response curve [41].

$$\left[\% \text{ Cell Viability} = \frac{(\text{Mean OD of Individual Sample})}{(\text{Mean OD of Control})}\right] * 100$$

3. Results and Discussion

3.1 Synthesis

3.1.1 N-((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1Hbenzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl) substituted benzamide derivatives (T16-T30)

(S)-1-amino-3-methyl-1,3,4,5-tetrahydro-2Hbenzo[d]azepin-2-one undergoes amidation with (tertbutoxycarbonyl)-L-alanine, gives tert-butyl ((S)-1-(((S)-3methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1-

yl)amino)-1-oxopropan-2-yl)carbamate (Inter-1), which is further undergoing deprotection gives (S)-2-amino-N-((S)-3methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1-

yl)propanamide (Inter-2) (Figure 1). Inter-2 on reaction with different substituted aromatic carboxylic acids and yields N-((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1H-

benzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl) substituted benzamide derivatives (T16-T30) with the formation of amide bond. Inter-2 undergoes nucleophilic addition reactions with different substituted benzoic acids and yields final derivatives T16-T30 after 12-16 hr reflux in THF solvent. Derivatives yielded 60% to 70% and were recrystallized with ethanol (Figure 2).

3.2 Spectral Interpretation of Synthesized Compounds

3.2.1 Tert-butyl ((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5tetrahydro-1H-benzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl)carbamate (Inter-1)

68% yield; ¹H NMR: δ 1.33-1.48 (12H, d), 1.43 (s)), 2.81-2.97 (2H, ddd), 3.08 (3H, s), 3.47-3.71 (2H, ddd), 4.47 (1H, q), 5.65 (1H, s), 7.06-7.31 (4H, td), 7.21 (ddd), 7.22 (ddd), 7.24 (ddd); MASS: 361.20 M/Z.

3.2.2(S)-2-amino-N-((S)-3-methyl-2-oxo-2,3,4,5-

tetrahydro-1H-benzo[d]azepin-1-yl) propanamide (Inter-2) 64.85% yield; ¹H NMR: δ 1.32 (3H, d), 2.81-2.97 (2H, ddd), 3.08 (3H, s), 3.47-3.71 (3H, ddd), 3.66, q), 5.69 (1H, s), 7.06-7.31 (4H, td); MASS: 261.15M/Z.

3.2.3 N-((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1Hbenzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl)substituted benzamide (T16-T30)

62-75% yield; ¹H NMR: δ 1.39 (3H, d), 2.81-2.97 (2H, ddd), 2.88 (ddd), 3.08 (3H, s), 3.47-3.71 (2H, ddd), 3.62 (ddd), 4.45 (1H, q), 5.72 (1H, s), 6.72 (2H, dd), 7.06-7.31 (4H, td), 7.21 (ddd), 7.22 (ddd), 7.24 (ddd), 7.40 (1H, t).

3.3 In Vitro Cytotoxicity Assay

The MTT assay was performed to screen the synthesized compounds on Vero and MDA MB 231 cells, and the results are shown in Table 8, Figure 3. Even at higher concentrations of synthesized derivatives for treatment, no cytotoxicity was observed in normal cells. Title compounds have exhibited low to high *in-vitro* anticancer activity with MDA MB 231 cells. Four of the synthesized compounds have shown good IC₅₀ values like T22 (15.105 μ g/ml), T24 (37.206 μ g/ml), T18 (41.200 μ g/ml), T20 (58.302 μ g/ml).





Figure 1: Synthetic scheme for the synthesis of N-((S)-1-(((S)-3-methyl-2-oxo-2,3,4,5-tetrahydro-1H-benzo[d]azepin-1-yl)amino)-1-oxopropan-2-yl)substituted benzamide derivatives (T16-T30)

S. No.	Chemicals and Solvents	M.Wt.	mM.	Eq.	Qty.	Unit
1	MC07JC1876	190	57.9	1.0	11	g
2	N-Boc Alanine	189	58.2	1.0	11	g
3	HOBt hydrate	153	64.05	1.1	9.8	g
4	EDC.HCL	191	64.0	1.1	12.3	g
5	N,N Diisopropyl ethyl amine	129	65.1	1.12	8.4	g
6	THF	-	-	-	200	mL

Table 2: Quantities of Chemicals and Solvents



Figure 2: Chemical Structures of synthesized compounds (T16-T30)



Figure 3: Cytotoxicity results of synthesized small molecules on both Vero and MDA MB 231 cell lines



Fig 4: Chen	nical structures	s of T18,	T20,	T22	and '	Т24
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S. No.	Chemicals and Solvents	M.Wt.	mM.	Eq.	Qty.	Unit
1	Inter-1	362	27.62	1.0	10	g
2	Dioxane	-	-	-	-	mL
3	HCl gas	-	-	-	-	

Table 3: Quantities of Chemicals and Solvents

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Table 4: Quantities of Chemicals and Solvents

S. No.	Chemicals and Solvents	M.Wt.	mM.	Eq.	Qty.	Unit
1	Inter-2	262	22.90	1.0	6	g
2	(S)-2-hydroxy-3-methyl butyric acid	118	22.88	1.0	2.7	g
3	HOBt hydrate	153	25.16	1.1	3.85	g
4	EDC.HC1	191	25.36	1.1	4.84	g
5	N,N-diisopropylethylamine	129	32.17	1.4	4.15	g
6	THF	-	-	-	60	mL

Table 5: R Group Substitutions of compounds synthesized (T16-T30)

S. No	Compound	Substitution (R)		
1	T16	Н		
2	T17	3,4-dihydroxy		
3	T18	3,4,5-trihydroxy		
4	T19	2-hydroxy		
5	T20	4-Hydroxy, 3,5-di Methoxy		
6	T21	2,6- diHydroxy		
7	T22	(Z)-3-(3,4-dihydroxyphenyl)acrylic acid		
8	T23	(Z)-3-(3-methoxy, 4-hydroxyphenyl)acrylic acid		
9	T24	(Z)-3-(phenyl)acrylic acid		
10	T25	(Z)-3-(4-hydroxyphenyl)acrylic acid		
11	T26	4-Nitro		
12	T27	4-Bromo		
13	T28	4-Chloro		
14	T29	4-Methyl		
15	T30	4-N,N-dimethyl		

Table 6: Physical Properties of synthesized compounds (T16-T30)

S. No	Compounds	Mol.	Mol.	Melting	**Rf
		Formula	weight	point (°C)	values
1	T16	$C_{21}H_{23}N_3O_3$	365.43	102-105	0.57
2	T17	$C_{21}H_{23}N_3O_5$	397.43	110-104	0.62
3	T18	$C_{21}H_{23}N_3O_6$	413.43	120-123	0.49
4	T19	$C_{21}H_{23}N_3O_4$	381.43	99-102	0.55
5	T20	$C_{23}H_{27}N_3O_6$	441.48	98-101	0.62
6	T21	$C_{21}H_{23}N_3O_5$	397.43	103-106	0.36
7	T22	$C_{24}H_{27}N_3O_5$	437.50	110-113	0.10
8	T23	$C_{23}H_{25}N_3O_3$	391.47	192-195	0.78
9	T24	$C_{23}H_{25}N_3O_4$	407.47	191-194	0.12
10	T25	$C_{21}H_{22}N_4O_5$	410.16	98-100	0.52
11	T26	$C_{21}H_{22}N_4O_5$	410.43	182-183	0.12
12	T27	$C_{21}H_{22}BrN_3O_3$	444.33	175-178	0.86
13	T28	$C_{21}H_{22}ClN_3O_3$	399.88	105-107	0.53
14	T29	$C_{22}H_{25}N_3O_3$	379.46	102-105	0.57
15	T30	$C_{23}H_{28}N_4O_3$	408.50	192-195	0.78

S. No	Compound	VERO (µg/ml)	MDA MB 231 (µg/ml)
1	T16	97	65
2	T17	90	115
3	T18	67	41
4	T19	152	72
5	T20	109	58
6	T21	124	98
7	T22	31	15
8	T23	296.74	148
9	T24	61	37
10	T25	611	115
11	T26	298	141
12	T27	98	64
13	T28	101	88
14	T29	88	72
15	T30	303.15	155
Standard	Raloxifene	15	6

Table 7: Cytotoxicity results of synthesized benzoazepinone derivatives

4. Conclusions

We have designed, synthesized and characterized a group of ER α antagonists that mimic the manner in which SERM and inserts into a cavity within ER α and inhibits its activity. A total of 15 compounds were synthesized by using a diverse scheme and the title compounds have exhibited low to high *in-vitro* anticancer activity with MDA MB 231 cells. Compared to the standard, the developed compounds T18, T20, T22 and T24 show considerable *in vitro* activity.

Finally, four compounds might be used as a lead molecule for future development into a therapeutically viable anti-ER positive breast cancer drug from the benzoazepinone derivatives family.

References

- Bandameedi R, Mohammed S, Soma H. A case study on National List of Essential Medicines (NLEM) in India and WHO EML 2015overview. *Pharm Regul Aff.* 2016;5:159.
- [2] H. Sung, H, J. Ferlay, R.L. Siegel, M. Laversanne, I. Soerjomataram, A. Jemal & F.Bray. (2021). Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA: A Cancer Journal for Clinicians.71, 209–249, doi:10.3322/CAAC.21660.
- [3] Globocon. (2018. Available online: http://cancerindia.org.in/globocan-2018-indiafactsheet/.
- [4] M. Clarke. (2006). Meta-analyses of adjuvant therapies for women with early breast cancer: The Early Breast Cancer Trialists' Collaborative Group overview. Annals of oncology. 17, doi:10.1093/annonc/mdl238.

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- [5] R.C. Coombes, L. Gibson, E. Hall, M. Emson & J. Bliss. (2003). Aromatase inhibitors as adjuvant therapies in patients with breast cancer. Journal of Steroid Biochemistry and Molecular Biology. 86, 309–311, doi:10.1016/S0960-0760(03)00372-8.
- [6] S. Jubie, A. Basheer, N. Yadav, A. Wadhwani & M. A. Azam. (2019). Synthesis and evaluation of coumate analogues as estrogen receptor inhibitors for breast cancer therapy. International journal of pharmaceutical sciences research. 10, 2218–2224, doi:10.26452/ijrps.v10i3.1455.
- [7] A.J. Begam, A. Basheer, S. Jubie & S.P. Dhanabal. (2017). An in-silico study of novel coumarin analogues as huamn estrogen receptor inhibitors. Journal of Pharmaceutical Sciences and Research. 9, 1953–1957.
- [8] A.J. Begam, S. Jubie & M.J. Nanjan. (2017). Estrogen receptor agonists/antagonists in breast cancer therapy: A critical review. Bioorganic Chemistry.71,257–274. doi:10.1016/j.bioorg.2017.02.011
 - doi:10.1016/j.bioorg.2017.02.011.
- [9] J. Selvaraj, J.B.A. John, N.M. Joghee, J. Antony, A. Wadhwani & J. Natarajan. (2019). Coumarin-Fatty Acid Conjugates as Potential ERα/AKT-1 Antagonists for ER Positive Breast Cancer. Anti-Cancer Agents in Medicinal Chemistry. 20, 437– 449, doi:10.2174/1871520619666191028104339.
- [10] S. Johnston. (2004). Fulvestrant and the sequential endocrine cascade for advanced breast cancer. British journal of cance. 90, S15–S18, doi:10.1038/sj.bjc.6601632.
- [11] W.M. ElShamy & D.M. Livingston. (2004). Identification of BRCA1-IRIS, a BRCA1 locus 812

product. Nature cell biology. 6, 954-967. doi:10.1038/ncb1171.

- [12] A. Cindy. Wilson, L. Ramos, R. Maria, Villaseñor, H. Karl. Anders & F. Michael. (1999). Press, Kathy Clarke, Beth Karlan, Jun-Jie Chen, Ralph Scully, David Livingston, Robert H. Zuch, Michael H. Kanter, Sylvan Cohen, F.J.C.& D.J.S. Localization of human BRCA1 and its loss in high-grade, noninherited breast carcinomas. Nature genetics. 21, 236-240.
- [13] C.X. Deng. (2006). BRCA1: Cell cycle checkpoint, genetic instability, DNA damage response and cancer evolution. Nucleic acids research. Special publication.34,1416-1426, doi:10.1093/nar/gkl010.
- [14] R.I. Yarden & M.Z. Papa. (2006). BRCA1 at the crossroad of multiple cellular pathways: Approaches for therapeutic interventions. Molecular cancer therapeutics. 5, 1396-1404, doi:10.1158/1535-7163.MCT-05-0471.
- [15] J. Dine & C.X. Deng. (2013). Mouse models of BRCA1 and their application to breast cancer research. Cancer metastasis reviews. 32, 25-37, doi:10.1007/s10555-012-9403-7.
- [16] N.E. Buckley & P.B. Mullan. (2012). BRCA1 -Conductor of the Breast Stem Cell Orchestra: The Role of BRCA1 in Mammary Gland Development and Identification of Cell of Origin of BRCA1 Mutant Breast Cancer. Stem Cell Reviews and Reports. 8, 982-993, doi:10.1007/s12015-012-9354-y.
- [17] C.X. Deng & R.H. Wang. (2003). Roles of BRCA1 in DNA damage repair: A link between development and cancer. Human molecular genetics. 12, doi:10.1093/hmg/ddg082.
- [18]L. Cao, W. Li, S. Kim, S.G. Brodie & C.X. Deng. (2003). Senescence, aging, and malignant transformation mediated by p53 in mice lacking the Brca1 full-length isoform. Genes and Development. 17, 201, doi:10.1101/GAD.1050003.
- [19] L. Cao, W. Li, S. Kim, S.G. Brodie & C.-X. Deng. (2003). Senescence, aging, and malignant transformation mediated by p53 in mice lacking the Brca1 full-length isoform. Genes & Development. 17. 201. https://doi.org/10.1101/GAD.1050003.
- [20] T. Miki, D.P. Bottaro, T.P. Fleming, C.L. Smith, W.H. Burgess, A.M.L. Chan & S.A. Aaronson. (1992). Determination of ligand-binding specificity by alternative splicing: Two distinct growth factor receptors encoded by a single gene. Proceedings of the National Academy of Sciences of the United States of America. 246-250. 89, https://doi.org/10.1073/PNAS.89.1.246.
- [21] D.F. Easton, D. Ford & D.T. Bishop. (1995). Breast and ovarian cancer incidence in BRCA1-mutation carriers. Breast Cancer Linkage Consortium. American Journal of Human Genetics. 56. 265.
- [22] J.P. Struewing, P. Hartge, S. Wacholder, S.M. Baker, M. Berlin, M. McAdams, M.M. Timmerman, L.C. Brody & M.A. Tucker. (1997). The Risk of Cancer Associated with Specific Mutations of BRCA1 and BRCA2 among Ashkenazi Jews. New England Journal of Medicine. 336, 1401-1408. https://doi.org/10.1056/nejm199705153362001.

[23] D. Ford, D.F. Easton, M. Stratton, S. Narod, D. Goldgar, P. Devilee, D.T. Bishop, B. Weber, G. Lenoir, J. Chang-Claude, H. Sobol, M.D. Teare, J. Struewing, A. Arason, S. Scherneck, J. Peto, T.R. Rebbeck, P. Tonin, S. Neuhausen, R. Barkardottir, J. Eyfjord, H. Lynch, B.A.J. Ponder, S.A. Gayther, J.M. Birch, A. Lindblom, D. Stoppa-Lyonnet, Y. Bignon, A. Borg, U. Hamann, N. Haites, R.J. Scott, C.M. Maugard, H. Vasen, S. Seitz, L.A. Cannon-Albright, A. Schofield & M. Zelada-Hedman. (1998). Genetic heterogeneity and penetrance analysis of the BRCA1 and BRCA2 genes in breast cancer families. American Journal of Human Genetics.62,676-689.

https://doi.org/10.1086/301749.

- [24] D.S.-E. Y. Miki & J. Swensen. (1994). A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1, Science. 266, 66-71.
- [25] T.F. Lane, C. Deng, A. Elson, M.S. Lyu, C.A. Kozak & P. Leder. (1995). Expression of Brca1 is associated with terminal differentiation of ectodermally and mesodermally derived tissues in mice. Genes & Development. 9, 2712-2722. https://doi.org/10.1101/GAD.9.21.2712.
- [26] J.A. Garson, P.R. Grant, U. Ayliffe, R.B. Ferns & R.S. Tedder. (2015). Real-time PCR quantitation of hepatitis B virus DNA using automated sample preparation and murine cytomegalovirus internal control. Journal of Virological Methods. 126, 207-213.

https://doi.org/10.1016/j.jviromet.2005.03.001.

- [27] D.G. Ginzinger. (2002). Gene quantification using real-time quantitative PCR: An emerging technology hits the mainstream. Experimental Hematology. 30. 503-512. https://doi.org/10.1016/S0301-472X(02)00806-8.
- [28] M.L. Wong & J.F. Medrano. (2018). Real-time PCR for mRNA quantitation. Biotechniques. Https://Doi.Org/10.2144/05391RV01. 39, 75-85. https://doi.org/10.2144/05391RV01.
- [29] S.A. Bustin. (2000). Absolute quantification of mrna using real-time reverse transcription polymerase chain reaction assays. Journal of Molecular 169-193. Endocrinology. 25. https://doi.org/10.1677/jme.0.0250169.
- [30] M.W. Pfaffl, A. Tichopad, C. Prgomet & T.P. Neuvians. (2004). Determination of stable housekeeping genes, differentially regulated target genes and sample integrity: BestKeeper - Excelbased tool using pair-wise correlations. Biotechnology Letters. 26:6. 26, 509-515. https://doi.org/10.1023/B:BILE.0000019559.84305 .47.
- [31] Naresh, P. Wadhwani, A. Jubie & Selvaraj. (2021). Dual Modulators of p53 and Cyclin D in ER Alpha Signaling by Albumin Nanovectors Bearing Zinc Chaperones for ER-positive Breast Cancer Therapy. Mini-Reviews in Medicinal Chemistry. 21,792-802. https://doi.org/10.2174/1389557520999201124212 347.
- [32] A. Jameera Begam, S. Jubie & M.J. Nanjan. (2017). Estrogen receptor agonists/antagonists in breast cancer therapy: A critical review. Bioorganic

Chemistry.71,257–274.

https://doi.org/10.1016/j.bioorg.2017.02.011.

[33] S. Jubie, S. Sundar, N. Yadav, P. Naresh, A. Wadhwani & J. Natarajan. (2019). A New Class of Coumate Benzimidazole Hybrids as BRCA 1 Mimetics Through Unconventional Binding Mode; Synthesis and Preliminary Cytotoxicity Screening. Current Computer-Aided Drug Design. 16, 786– 801.

https://doi.org/10.2174/1573409916666191231102 046.

- [34] Begam, A.J.J Basheer, K.A. Jubie, S. Jupudi, S. Azam, M.A. Dhanabal & Palanisamy. (2019). A New Class of Pure Estrogen Alpha Receptor Antagonists; Design, Synthesis and in-vitro Screening. Letters in Drug Design & Discovery.16,66–81. https://doi.org/10.2174/1570180815666180327124 634.
- [35] P. S. Sundar, P. Naresh, P. Natarajan, J.A. Wadhwani & S. Jubie. (2021). Potential Coumarin Thiosemicarbazone Hybrids as BRCA-1 Mimetics for ER Positive Breast Cancer Therapy: An In-silico Approach. Journal of Medical Pharmaceutical and Allied Sciences. 10, 3484–3493. https://doi.org/10.22270/jmpas.V10I4.1383.
- [36] P. Thangavelu & J. Selvaraj. (2020). QSAR Approach to Improve the Estrogen Inhibiting Activity of Coumate Analogs: A Strategy to Treat the Breast Cancer.
- [37] Miki, Y. Swensen, J. Shattuck-Eidens, D. Futreal, P.A. Harshman, K. Tavtigian, S. Liu, Q. Cochran, C. Bennett, L. M. Ding & Wei. (1994). A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1. Science. (New York, N.Y.). 266, 66–71. https://doi.org/10.1126/SCIENCE.7545954.
- [38] J. Taylor, M. Tymboura, P.E. Pace, R.P. A'Hern, A.J. Desai, S. Shousha, R.C. Coombes & S. Ali. (1998). An important role for BRCA1 in breast cancer progression is indicated by its loss in a large proportion of non-familial breast cancers. International Journal of Cancer. 79. 334–342. https://doi.org/10.1002/(SICI)1097-0215(19980821)79:4<334::AID-IJC5>3.0.CO;2-W.

[39] M. Esteller. (2000). Promoter Hypermethylation and BRCA1 Inactivation in Sporadic Breast and Ovarian Tumors. Journal of the National Cancer Institute.92,564–569. https://doi.org/10.1093/jnci/92.7.564.