

International Journal of Chemical and Biochemical Sciences (ISSN 2226-9614)

Journal Home page: www.iscientific.org/Journal.html



© International Scientific Organization

Unleashing the Power of *Colchicum luteum* Leaf Powder for Hexavalent

Chromium [Cr(VI)] Removal in Water - A Kinetic Study

Ujjwal Kumar¹, Usha Sharma² Ashok K. Jha^{3*}, Pallavi Kumari³, Shailesh Kumar³

¹P.G. Department of Biotechnology, ²Department of Chemistry G.B. College Naugachia ³University Department of Chemistry, T.M. Bhagalpur University, Bhagalpur – 812 007, India

Abstract

This research paper investigates the efficacy of a biosorbent derived from leaf powder of *Colchicum luteum* for the mitigation of hexavalent chromium Cr(VI) from aqueous solution. The study aims to explore the potential of utilizing natural materials as a cost-effective and eco-friendly approach for heavy metal remediation. The biosorbent was prepared by processing the dried leaves of *C. luteum* into a fine powder. Batch experiments were conducted to evaluate the biosorption capacity of the prepared biosorbent for Cr(VI) removal. The effects of various parameters, including initial Cr(VI) concentration, contact time, temperature, concentration of solution and biosorbent dosage, were investigated to understand their influence on the biosorption process. The results of the study indicated that the biosorption of Cr(VI) onto *C. luteum* leaf powder followed the Freundlich, Langmuir, and Elovich isotherm models. The biosorption efficiency decreased with increasing biosorbent dose, suggesting the involvement of intraparticle diffusion. Further, the contact time and initial ion concentration influenced the biosorption process, with the maximum biosorption observed at 3 ppm Cr(VI) concentration.

Keywords: Colchicum luteum, Hexavalent chromium [Cr(VI)], Biosoption, Isotherm models.

Full length article *Corresponding Author, e-mail: ashokjha39@gmail.com

1. Introduction

Sura-ranjan (Colchicum leuteum) has various uses in Ayurvedic medicine such as liver disorder, spleen and skin problems, gout rheumatic disorder and leukaemia [1-2]. The chemical constituents are colchicine, demecolcine, colchifoline, N-formyl colchicine and corregerine are important Phytochemicals present in different species [3]. Medicinal plants have gained importance all over the world because of its long term relief against many diseases and lesser side effect [4]. But use of raw medicinal plants growing on a soil containing heavy metals can cause adverse health effects [5]. Generally, Pb, Cd, Cr are accumulated by the medicinal plants [6-8]. Detailed information on the medicinal properties, phytoremediation of Cr(VI), and specific information on C. luteum has been reported [9]. In view of these problems, the heavy metal uptake such as Cr(VI) by Colchicum Leuteum was studied and the data analysed for different isotherm [9-11]. Due to rapid industrialization and urbanization, aquatic environments are under imminent danger and facing a catastrophe. Hexavalent chromium Cr(VI), excessively released from various industries, has become a global concern [12]. Cr(VI) receives significant attention due to its widespread industrial applications and stability in the natural environment [13]. In some areas of the Gangetic basin in India, Cr(VI) naturally occurs in groundwater aquifers, leading to groundwater contamination.

The World Health Organization (WHO) has set a permissible concentration of Cr(VI) in water bodies at 0.05 ppm. Therefore, it is essential to decontaminate Cr(VI) from industrial wastewater before disposal and its return to the natural environment [14-15]. Numerous conventional methods have been developed and successfully implemented for Cr(VI) mitigation in water bodies. However, these methods are often cost-effective and technologically complicated, making them impractical for remote villages and distant locations. In recent years, several methods utilizing plant-based bio-sorbents have been developed for the treatment of heavy metals. Plant materials are advantageous as they are inexpensive, readily available, and can be applied without the need for external energy sources [16]. Various medicinal plants and herbs have been identified as effective agents for decontaminating Cr(VI) from aqueous solutions [17-18]. After extracting the medicinally important bioactive compounds, the waste parts of medicinal plants can be utilized as biosorbents for Cr(VI) mitigation [19]. One such plant of interest is C. luteum, also known as Suranjan. The objective of this work is to evaluate the biosorption capacities and kinetics of Cr(VI) using Langmuir, Freundlich, and Elovich isotherms.

2. Material and methods

2.1. Preparation of biosorbent from leaf biomass of C. luteum.

Healthy and green leaves of *C. luteum* were selected for biosorption studies of Cr(VI). Green leaves were cut into small pieces and also washed five times with running tap water and again thrice with distilled water. After that, the sample was air dried and cut into more small pieces again, put in an oven at 60 °C for 8 h (hour). After that, leave biomass was crushed in a grinder up to 200 mesh and again dried for 4 h at 60°C. Now biosorbent was prepared for batch operations.

2.2. Preparation of reagents

2.828 g of potassium dichromate $(K_2Cr_2O_7)$ is dissolved in 1000 ml deionized water to get 1000 ppm Cr(VI) solution. The working solutions were obtained by serial dilution of the stock solution to the appropriate volume and 1, 2, and 3 ppm solutions were prepared. Murasige and Skooge basal salt solution (Himedia) was also added to the solution for nutrient support to plants.

2.3. Batch biosorption experiment of Cr(VI) by leaf biomass of C. luteum

The biosorbent properties of leaf biomass of C. luteum on Cr(VI) ions were investigated in batch operation at room temperature (300 K). Cr(VI) removal treatment was carried out in a series of conical flasks (250 mL) for a known concentration of Cr(VI) i.e. 1, 2, and 3 ppm of 100 ml aqueous solution, taken separately in 250 mL stoppered conical flask. Initial pH was adjusted by 1N NaOH and HCl at 6.5 - 7.0 (Tab. 1-3). Different masses of biosorbent dose were added separately for all above-mentioned concentrations for different time intervals i.e. 1g biomass for 1, 2, and 3 h separately as well as different weights of biomass (1, 2, 3, and 4 g) biomass doses for 1 h time period separately. Apart from these blank, aqueous solutions were also measured (without biomass) as control. After completion of treatment time, biosorbent was separated by filter paper and the residual solution of Cr(VI) was analyzed by UV double beam spectrophotometer (Pharo300) (Tab. 1-3). Cr(VI) was estimated by Spectroquant Chromate test kit (Merck). Total Cr(VI) present in the sample was determined after pretreatment of the sample by using this kit. In this procedure dry test tube is taken and 1 level microspoon (assembled in the cap of the bottle) Cr-1 reagent is added then 6 drop of reagent Cr-2 is added, provided in the kit. After that it is shaken properly until the reagent is completely dissolved then add 5 mL residual sample. It is left for 1 min at room temperature and measured in the spectrophotometer by balancing bar code for set up wave length. The concentration of Cr(VI) was also estimated by diphenyl carbazide method from UV double beam spectrophotometer systronics 2203 model. The values of concentration are in agreement with each other. The amount of Cr(VI) sorbed per unit mass adsorbent was evaluated by using mass balance equation (I) and the removal percentage was also determined according to the given equations (II). The adsorption amount per unit mass of adsorbent has been calculated by the equation given below. $q_t = (C_0 - C_t) V/W$ (I) where,

 q_t is (mg/g) the amount of As (III) and Cr(VI) absorbed after time t in minutes. C₀ represents the initial concentration and C_t final concentration. V is the volume of As (III) and Cr(VI) solution (mL) and W is the weight of bio sorbent (g). The removal percentage (%) of Cr(VI) ions from the aqueous solution after biosorption was calculated by applying the following equation.

Removal (%) = $(C_0 - C_t)/C_t \times 100$ where,

 C_0 and C_t were the initial and final concentrations respectively for Cr(VI) solution after the biosorption process.

(II)

2.4. Sorption isotherm models

Modelling of sorption isotherm data is important for predicting and comparing the sorption performance of the biosorbent. Therefore, the equilibrium data fitted using different isotherm models, namely Langmuir, Freundlich and Elovich. Sorption isotherms were performed by leave biomass of *C. luteum* in above mentioned batch condition and isotherm models were validated by their respective linear regression coefficient (\mathbb{R}^2) as given in Fig 1- 10. To estimate the type of interaction involved, adsorption mechanism is derived by adsorption isotherms. Analysis of equilibrium sorption data of Cr(VI) onto biosorbent of C. luteum fitted to *Freundlich isotherm, Langmuir isotherm* and Elovich isotherm models, whose linear equations are given (III), (IV) and (V), respectively.

$$\log q_t = \log K_F + \frac{1}{n} \log C_t \tag{III}$$

$$\frac{1}{q_t} = \frac{1}{K_L q_m} \left(\frac{1}{C_t}\right) + \frac{1}{q_m}$$
 (IV)

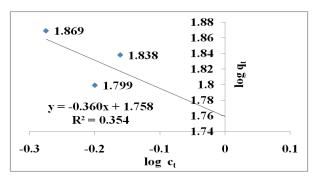
$$\ln(\frac{q_t}{c_t}) = \ln K_e q_m - \frac{q_t}{q_m}$$
(V)

In the above equations $q_e =$ amount of adsorbate (mg) adsorbed per unit mass (g) of adsorbent, $C_e =$ the equilibrium concentration (mgL⁻¹) of the adsorbate in the solutions. The applicability of Freundlich, Langmuir and Elovich isotherm analysis were determined by linear regression coefficient (R² Value) using Freundlich plot (log q_t versus log C_t), Langmuir plot ($\frac{1}{q_t}$ versus $\frac{1}{C_t}$) and Elovich plot (ln ($\frac{q_t}{C_t}$) versus q_t) applied on batch equilibrium data at room temperature 308 K.

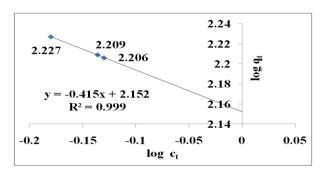
3. Results and discussion

3.1. Isotherm analysis of biosorption by leave biomass

The type of adsorption mechanism involved in biosorption processes is determined by the adsorption isotherm model. Analysis of equilibrium sorption data of Cr(VI) on biosorbent dose of *C. luteum* fitted to Freundlich isotherm. Batch experiments for removal of Cr(VI) by leaf biomass of *C. luteum* from fixed weight (1g) of biosorbent dose with 1, 2 and 3 h time treatment separately, plots show linear regression coefficient (R²) of Freundlich isotherm as 0.354, 0.999 and 0.999 (Fig. 1) for 1, 2 and 3 ppm initial concentration dose respectively. These results confirm that 1 ppm initial concentration does not obey the Fruendlich model while the other two initial ion batches i.e. 2 ppm and 3 ppm exactly fitted on Freundlich model. In further studies, different weights of powdered biomass (1g, 2g, 3g, and 4 g) for fixed time intervals (1 h) were taken separately.







B

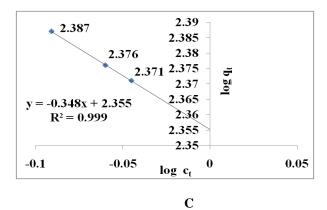


Figure 1: Plot of Freundlich isotherm of Cr(VI) biosorption by fixed weight biomass of *C. luteum* (1g) with different time interval (1h, 2h and 3h). Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

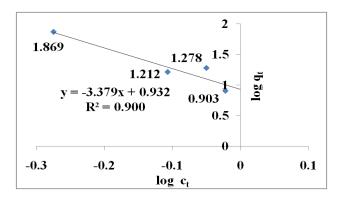
IJCBS, 24(5) (2023): 167-182

Biomass (g) with		1 ppm Cr(VI) initial ion concentration										
		Ct	qt	log	log	R ²	1/C _t	1/q _t	R ²	In	R ²	
time	C0			Ct	qt	(Freundlich)			(Langmuir)	(q _t /C _t)	(Elovich)	
interval												
(h)												
1g for 1h		0.53	74	-	1.869		1.886	0.013		4.864		
				0.275		0.354			1.0		0.817	
1g for 2h	1.27	0.64	63	-	1.799		1.562	0.015		2.286		
	1.27			0.200								
1g for 3h		0.58	69	-	1.838		1.724	0.014		4.778		
				0.161								
1g for 1h		0.53	74	-	1.869		1.886	0.013		4.864		
				0.275							0.944	
2g for 1h		0.89	19	-	1.278	0.900	1.123	0.052	0.652	3.060		
				0.050		0.900			0.052			
3g for 1h		0.78	16.33	-	1.212		1.282	0.061		3.0411		
				0.107								
4g for 1h		0.95	8	-	0.903		1.052	0.125		2.130		
				0.022								

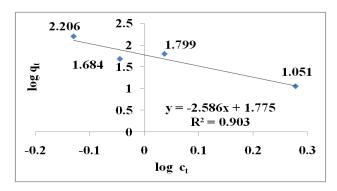
Table 1: Langmuir, Freundlich and Elovich isotherm parameters for adsorption of Cr(VI) onto leave surface of *C. luteum* at room temperature and 1 ppm initial concentration

Table 2: Langmuir, Freundlich and Elovich isotherm parameters for adsorption of Cr(VI) onto leave surface of *C. luteum* at room temperature and 2 ppm initial concentration

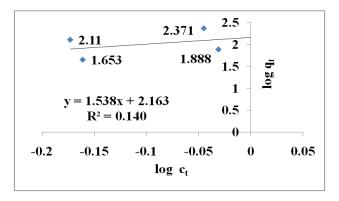
Biomass (g) with time interval (h)		2 ppm Cr(VI) initial ion concentration									
	C0	Ct	qt	log C _t	log q _t	R² (Freund lich)	1/C _t	1/q _t	R² (Langmu ir)	In (q _t /C _t)	R ² (Elovich)
1g for 1h		0.74	161	-0.130	2.206		1.351	0.0062		5.382	
1g for 2h	2.35	0.66	169	-0.180	2.227	0.999	1.515	0.0059	0.995	5.545	1.0
1g for 3h		0.73	162	-0.136	2.209		1.396	0.0061		5.402	-
1g for 1h		0.74	161	-0.130	2.206		1.351	0.0062		5.382	
2g for 1h		1.09	63	0.037	1.799	0.903	0.917	0.0161	0.823	4.056	1
3g for 1h		0.90	48.33	-0.045	1.684		1.111	0.0206		3.983	0.805
4g for 1h		1.90	11.25	-0.278	1.051		0.526	0.0888		1.77	





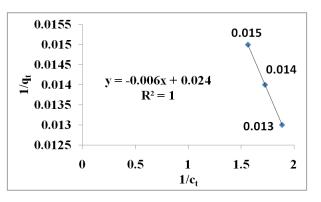




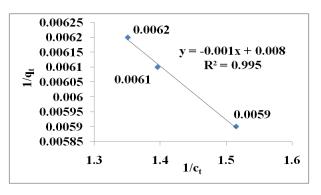


(**C**)

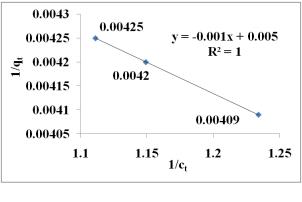
Figure 2: Plot of Freundlich isotherm of Cr(VI) biosorption by different weight biomass of *C. luteum* (1g, 2g, 3g and 4g) with uniform time interval (1h). Initial Cr(VI) concentration of 1 ppm (**A**), Initial Cr(VI) concentration of 2 ppm (**B**) and initial Cr(VI) concentration of 3 ppm (**C**),





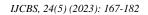


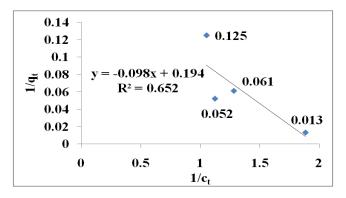




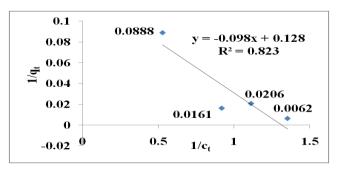
(**C**)

Figure 3: Plot of Langmuir isotherm of Cr(VI) biosorption by fixed weight biomass of *C. luteum* (1g) with different time interval (1h, 2h and 3h). Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

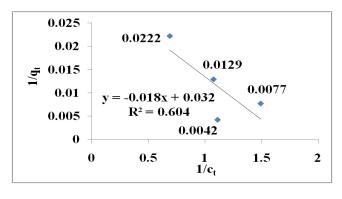








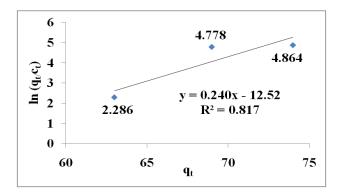




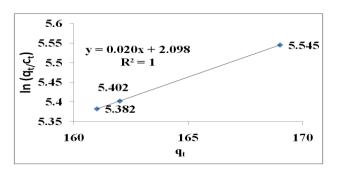
(**C**)

Figure 4: Plot of Langmuir isotherm of Cr(VI) biosorption by different weight biomass of *C. luteum* (1g, 2g, 3g and 4g) with uniform time interval (1h). Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

IJCBS, 24(5) (2023): 167-182









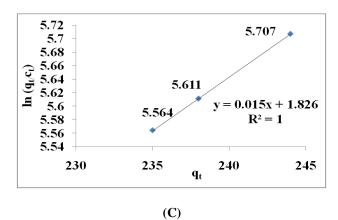
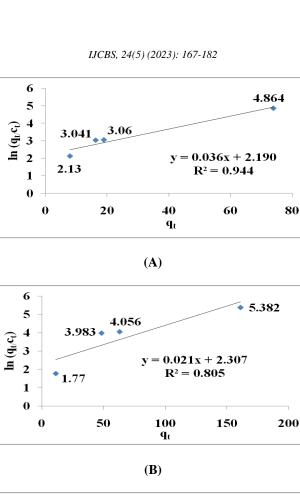
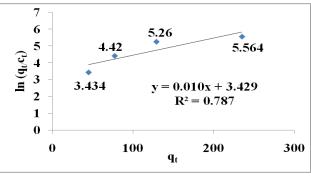


Figure 5: Plot of Elovich isotherm of Cr(VI) biosorption by fixed weight biomass of *C. luteum* (1g) with different time interval (1h, 2h and 3h). Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

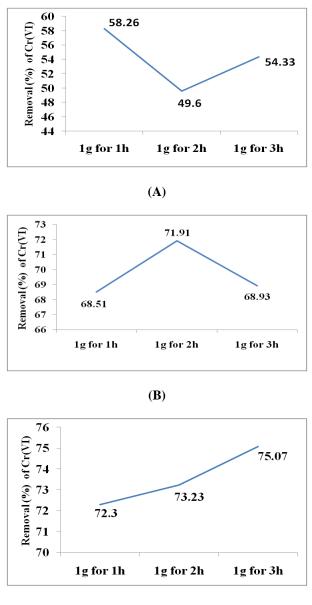




(**C**)

Figure 6: Plots of Elovich isotherm of Cr(VI) biosorption by different weight biomass of *C. luteum* (1g, 2g, 3g and 4g) with uniform time interval (1h). Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

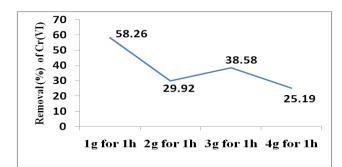
IJCBS, 24(5) (2023): 167-182



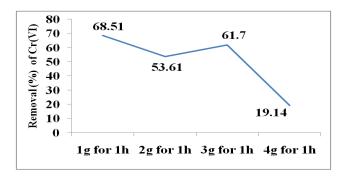
(C)

Figure 7: Graphs shows removal precentage (%) of Cr(VI) after biosorption by fixed biomass of *C. luteum* (1g) of sorbent dose with different time interval (1h, 2h and 3h) at different initial As(III) and Cr(VI) concentration. Initial Cr(VI) concentration of 1 ppm (A), initial Cr(VI) concentration of 2 ppm (B), and initial Cr(VI) concentration of 3 ppm (C).

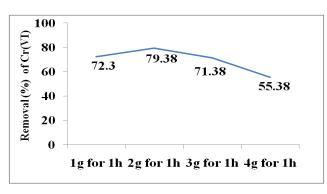
IJCBS, 24(5) (2023): 167-182



(A)







(C)

Figure 8: Graphs shows removal precentage (%) of Cr(VI) after biosorption by Different biomass (*C. luteum*) (1g, 2g, 3g and 4g) of sorbent dose with fixed time (1h) interval at different Initial Cr(VI) concentration. Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

IJCBS, 24(5) (2023): 167-182

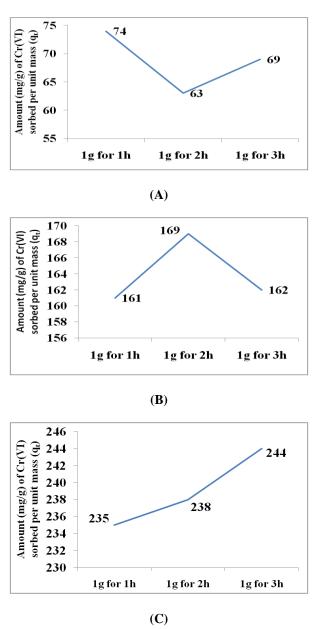


Figure 9: Amount of Cr(VI) sorbed per unit mass (qt) by fixed biomass (*C. luteum*) (1g) of sorbent dose with different time interval (1h, 2h and 3h) at different initial Cr(VI) concentration. Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**), and initial Cr(VI) concentration of 3 ppm (**C**).

IJCBS, 24(5) (2023): 167-182

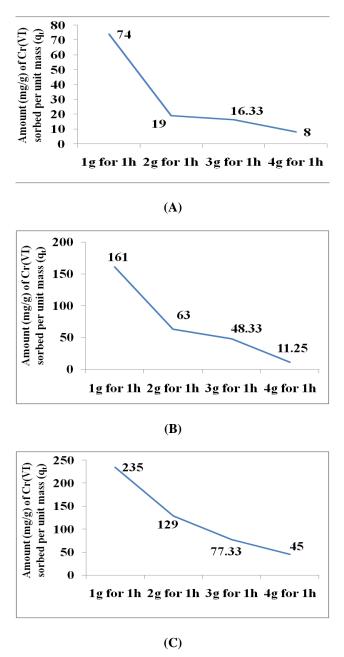


Figure 10: Amount of Cr(VI) sorbed per unit mass (q_t) by different biomass (*C. luteum*) (1g, 2g, 3g and 4g) of sorbent dose with uniform time interval (1h) at different initial Cr(VI) concentration. Initial Cr(VI) concentration of 1 ppm (**A**), initial Cr(VI) concentration of 2 ppm (**B**) and initial Cr(VI) concentration of 3 ppm (**C**).

Biomass (g) with		3 ppm Cr(VI) initial ion concentration									
		Ct	qt	log C _t	log q _t	R ²	1/C _t	1/q _t	R ²	In	R ²
time	CO					(Freund			(Langm	(q _t /C _t)	(Elovich)
interval						lich)			uir)		
(h)											
1g for 1h		0.90	235	-0.045	2.371		1.111	0.00425		5.564	
1g for 2h		0.87	238	-0.060	2.376	0.999	1.149	0.00420	1.0	5.611	1.0
1g for 3h	3.25	0.81	244	-0.091	2.387	-	1.234	0.00409		5.707	
1g for 1h		0.90	235	-0.045	2.371		1.111	0.00425		5.564	
2g for 1h		0.67	129	-0.173	2.110	1	1.492	0.0077	0.604	5.260	-
3g for 1h		0.93	77.33	-0.031	1.888	0.140	1.075	0.0129		4.420	0.787
4g for 1h		1.45	45	-0.161	1.653		1.689	0.0222	1	3.434	

Table 3: Langmuir, Freundlich and Elovich isotherm parameters for adsorption of Cr(VI) onto leave surface of *C. luteum* at room temperature and 3 ppm initial concentration.

 R^2 values of Freundlich isotherm were 0.900, 0.903, and 0.140 (Fig. 2) for 1, 2 and 3 ppm respectively. These data were perfect for 1 and 2 ppm concentrations in Freundlich plots whereas at 3 ppm it does not apply. Langmuir isotherm application for estimation of biosorption mechanism in all three initial ion concentrations of Cr(VI) was studied. The biosorption of Cr(VI) in the batch operation of fixed biomass (1 g) for different time intervals i.e. 1 h, 2 h, and 3 h was studied. Langmuir plots illustrated that R² values are 1.0, 0.995, and 1 for 1,2 and 3 ppm ion concentration respectively (Fig. 3). These plots show that Langmuir isotherm is the best fit. In another set where, different weights of biomass (1 g, 2 g, 3 g, and 4 g) were treated for the same time duration (1 h) separately, their Langmuir plots showed that their R² values were 0.652, 0.823 and 0.604 for 1 ppm, 2 ppm and 3 ppm respectively (Fig. 4). This analysis states that Langmuir model is the best fit on the biosorption studies when treatment time varied with a fixed weight of biomass. An illustration of the Elovich model is also applied to this study. In batch experiments of fixed biomass (1 g) with ascending time treatment (1h, 2 h and 3 h), plots show 0.817, 1.0 and 1.0 (Fig. 5) R² data respectively for 1 ppm, 2 ppm and 3 ppm ion concentrations. At batch of 1 ppm Elovich model is overall a good fit while at 2 ppm and 3 ppm it shows maximum R^2 value i.e. a fit of this model. These results established that Cr(VI) adsorption implies multilayer sorption and adsorption sites increase during biosorption. Another set with different ranges of biosorption dose (1 g, 2 g, 3 g and 4 g) with fixed time treatment (1h) shows 0.994, 0.805 and 0.787 (Fig. 6) for R^2 data of 1 ppm, 2 ppm and 3 ppm respectively, show overall a good fit. Elovich model is completely suitable for all sets of these experiments.

3.2. Effect of adsorbent dose in biosorption

The batch sorption experiment was carried out by four doses (1, 2, 3 and 4 g) in 100 ml Cr(VI) containing medium for 1 h treatment with initial ion concentrations of 1, 2 and 3 ppm (Tab. 1-3). Biosorption of Cr(VI) decreased when the adsorption dose is increased up to 4 g (Fig. 8). This trend may be due to intraparticle diffusion whereas the amount of Cr(VI) sorbed per unit biomass also show decreasing trend (Fig. 10) in all three ion concentrations. Further, fixed biomass (1 g) was treated for 1 h, 2 h and 3 h treatment, Cr(VI) ion sorbed per unit mass only showed an increase with 3 ppm (Fig. 9). Amount of sorbed ion on per unit mass was illustrated (Fig. 9, 10). Biosorption experiment with 1 g biomass for 1 h, 2 h and 3 h at 1 ppm, 2 ppm and 3 ppm initial Cr(VI) concentration respectively shows decreasing trends with respect to increasing time (Fig. 10) whereas, further study with different weight of biomass (1 g, 2 g, 3 g and 4 g) for 1h with 1 ppm, 2 ppm and 3 ppm initial Cr(VI) concentration. At 1 ppm and 2 ppm set, it also follows the decreasing rate of qt while at 3 ppm only shows increasing trends (Fig. 9).

Effect of contact time and ion concentration on biosorption

It was observed that the maximum biosorption took place in 3 ppm (Fig. 7, 8) ion concentration.

In different biosorbent doses (1, 2, 3 and 4 g) at 1 h time treatment for three different Cr(VI) concentration of 1 ppm, 2 ppm and 3 ppm, samples started showing desorption of Cr(VI) with almost every biosorption (Fig. 8). Further, 1 g biomass separately in 100 ml aqueous medium at 1 h, 2 h and 3 h time treatment, it was observed that effect of intraparticle diffusion had been found in 1 ppm and 2 ppm Cr(VI) ion

concentration whereas in 3 ppm ion concentration rate of biosorption increased with their contact time (Fig. 7).

The uptake of Cr(VI) by *Colchium leuteum* takes place either through bonding of ions to the surface complexation and electrostatic interaction [20-24].

4. Conclusions

Freundlich isotherm was best fitted on fixed leave biomass (1 g) experiment with 2 ppm and 3 ppm initial Cr(VI) concentration. On other hand Langmuir isotherm was completely fitted on all initial concentration of batches of fixed biomass set. Elovich isotherm was a fit for 1 ppm and 2 ppm batch of fixed biomass set whereas it only fitted on 1 ppm initial ion concentration from different weight of biomass set. In biosorption experiment of Cr(VI) by leave biomass of C. luteum, a maximum removal percent of 79.38 was observed. Thus the biosorption capacity varied with the initial concentration of Cr(VI) and the amount of biosorbent used. It was observed that the biosorption efficiency decreased with increasing biosorbent dose. The maximum biosorption was achieved at 3 ppm Cr(VI) concentration. Plant-based biosorbents offer a cost-effective and environmentally friendly alternative for heavy metal remediation. The uptake of Cr(VI) by Colchium leutium revealed that analysis of heavy metals should be done before consumption and contamination soil sites must be excluded for growing medicinal plants and drug preparation

References

- A. Husek, N. S.Pinar, P. Sedmera, F. Voegelein, I. Valka & V. Šimanek. (1990). Alkaloids and phenolics of Colchicum turcicum. Phytochemistry, 29(9): 3058-3060.
- [2] U. Kumar, AK Jha, RS Singh. (2021). Understanding the Bioaccumulation and Biosorption of Arsenic [As(III)] in Plants and Biotechnological Approaches for Its Bioremediation, Arsenic Toxicity: Challenges and Solutions. (Editor: Nitish Kumar). 363-389, ISBN: 978-981-33-6067-9, Springer Nature, Singapore
- G. Gulsoy-Toplan, F. Goger, A. Yildiz-Pekoz, S. Gibbons, G. Sariyar, A. Mat. (2018). Chemical Constituents of the different parts of Colchicum micranthum and C. chalcedonicum and their cytotoxic activities. Natural Product Communications, 13(5), https://doi.org/10.1177/1934578X1801300506
- [4] A.Davoodi, M. Azadbakht, S.J. Hosseinimehr, S.Emami, M. Azadbakht. (2021). Phytochemical profiles, physicochemical analysis, and biological activity of three Colchicum species. Jundishapur Journal of Natural Pharmaceutical Products. 2021 ;16(2): e98868, doi.org/10.5812/jjnpp.98868
- [5] A.V.Pethkar, R. P. Gaikaiwari and K. M. Paknikar. (2001). Biosorptive removal of contaminating heavy metals from plant extracts of medicinal value. Current Science, 2001;80,9: 1216-1219.
- [6] A.Kulhari, A.Sheorayan, S.Bajar, S. Sarkar, A. Chaudhury, R, K. Kalia. (2013). Investigation of heavy metals in frequently utilized medicinal plants collected from environmentally diverse locations of

north western India. Springer Plus. 2(1):1-9.https://doi.org/10.1186/2193-1801-2-676

- U. Kumar, Singh, R. S. Mandal, J. Nayak, A. K. Jha, (2022). Removal of As (III) and Cr (VI) from aqueous solutions by Bixaorellana leaf biosorbent and As (III) removal using bacterial isolates from heavy metal contaminated site. Journal of the Indian Chemical Society. 99(5); 100334. https://doi.org/10.1016/j.jics.2021.100334
- [8] Ashok K. Jha, U. Kumar, and Y. C. Gupta. (2015). Biosorption of heavy metals by aquatic weeds. Chemical Science Review and Letters. 4: 827-834.
- [9] Ashok K. Jha, and Ujjwal Kumar. (2017). Studies on removal of heavy metals by cymbopogonflexuosus." International Journal of Agriculture, Environment and Biotechnology. 10, 1 ;89-92. http://dx.doi.org/10.5958/2230-732X.2017.00017.1
- [10] U. Kumar, A.K. Jha, S. Kumar, and S. Verma. (2022). First report on bioaccumulation kinetics of Chromium (VI) and Malachite green by Colchicum luteum from aqueous medium. International Journal of Chemical and Biochemical Sciences. 21;139-147
- [11] S. Ambika, M. Kumar, L. Pisharody, M. Malhotra, G. Kumar, V. Sreedharan, L. Singh, P. V. Nidheesh, and A. Bhatnagar. (2022). Modified biochar as a green adsorbent for removal of hexavalent chromium from various environmental matrices: mechanisms method sandprospects. Chemical Engineering Journal. 439, 135716. https://doi.org/10.1039/C6RA07460J
- [12] Y. Li, J. Liu, Q. Yuan, H. Tang, F. Yu, X. Lv. (2016). A green adsorbent derived from banana peel for highly effective removal of heavy metal ions from water. RSC Advances. 6(51):450418, https://doi.org/10.1039/C6RA07460J
- P. Singh, N. Itankar, and Y. Patil. (2021). Biomanagement of hexavalent chromium: Current trends and promising perspectives. Journal of Environmental Management 279 111547.https://doi.org/10.1016/j.jenvman.111547
- [14] S.H. Abbas, H. Salman, I. M. Ismail, T. Mostafa, and H. A. Sulaymon. (2014). Biosorption of heavy metals: a review. Journal of Chemical Science and Technology. 3(4) 74-102.
- [15] B. Saha, and C. Orvig. (2010). Biosorbents for hexavalent chromium from industrial and municipal effluents, Co-ordination Chemistry Reviews 254, 2959-2972, doi.org/10.1016/j.ccr.2010.06.005
- [16] S. Mitra, A. Sarkar, and S. Sen. (2007). Removal of Chromium from industrial effluents using nanotechnology; a review, Nanotechnology for Environmental Engineering 2,1-14, doi.org/10.1007/S 41204-017-0022-y
- [17] D. Park, Yeoung –Sang Yun, and J.M. Park. (2006). Comment on the removal mechanism of hexavalent chromium by biomaterials or biomaterials-based activated carbons, Ind.Eng.Chem.Res.45,7,2405-2407
- [18] S. Prasad, K. K. Yadav, S. Kumar, N. Gupta, M.S. M. Cabral-Pinto, S. Rezania, N. Radwan, and J. Alam. (2021). Chromium contamination and effect

on environmental health and its remediation: A sustainable approaches." Journal of Environmental Management 285 112174.https://doi.org/10.1016/j.jenvman.2021.112 174

- C. Cervantes., J. Campos-García, S. Devars, F. Gutiérrez-Corona, H. Loza-Tavera, J. Carlos Torres-Guzmán, and R. Moreno-Sánchez. (2001). Interactions of chromium with microorganisms and plants." FEMS microbiology reviews 25, no. 3 335-347.https://doi.org/10.1111/j.1574-6976.2001.tb00581.x
- [20] U. Zulfiqar, FasihUllah Haider, M. Ahmad, S. Hussain, M.F. Maqsood, M. Ishfaq, B. Shahzad et al. (2023). Chromium toxicity, speciation, and remediation strategies in soil-plant interface: A critical review. Frontiers in Plant Science 13 1081624.https://doi.org/10.3389/fpls.2022.1081624
- [21] Q. Li, X. Zhuang, G. Zhou, Z. Yang, T. Yang, H. Xiao, T. Xu, and W. Wang. (2023). Efficient removal of Cr (VI) from wastewater by ZnO-polyacrylic acid/cellulose fiber/polyethylene glycol hydrogel: Synergistic effect of adsorption and photocatalytic reduction. Journal of Environmental Chemical

Engineering 110390.https://doi.org/10.1016/j.jece.2 023.110390

- [22] Gao-Jie Jiao, M. Jiliang, L.I. Yancong, J. Dongnv, Z. Jinghui, and S. Runcang. (2022). Removed heavy metal ions from wastewater reuse for chemiluminescence: Successive application of lignin-based composite hydrogels. Journal of Hazardous Materials 2022: 421 (126722.https://doi.org/10.1016/j.jhazmat.2021.126 722
- [23] A. T. Hoang, S. Nižetić, C. K. Cheng R Luque, S. Thomas, TL Banh, XP Nguyen. (2022). Heavy metal removal by biomass-derived carbon nanotubes as a greener environmental remediation: A comprehensive review. Chemosphere; 287:131959.https://doi.org/10.1016/j.chemosphere. 2021.131959
- [24] U. Kumar, A. K. Jha, R. Thakur. (2021). Bioremediation trends on mitigation of As(III), Cr(VI) and organic dyes from aqueous medium using plant and microbial biomass: An overview. International Journal of Bioresource and Stress Management. 12(6), 621-637.